Physicochemical Characterization of *Hymenopterasphecidae* (mud-wasp) nest

Adie P.A.¹, Kukwa D.T.^{1*}, Ikyereve R.E.² and Kungur P.D.¹

¹Department of Chemistry, Benue State University, Makurdi, NIGERIA ²Department of Chemistry, Loughborough University, Loughborough, Leicestershire, UNITED KINGDOM

Available online at: www.isca.in, www.isca.me

Received 17th June 2013, revised 8th July 2013, accepted 19th August 2013

Abstract

Hymenoptera sphecidae (mud-wasp) nests were harvested and evaluated for their physicochemical properties. Physicochemical characterization of mud-wasp nest gave the following values: pH 6.54; abrasion resistance, 95.10%; bulk density, 1.27 (g/mL); total surface charge, 1.348 (mmol H^+ eq/g); iodine adsorption number (IAN), 0.1656 (mmol/g); total surface area, 6.335 (g/mgI₂M): clay, 10.96%, silt, 15.52%; sand, 73.52%. Spectrophotometric analysis of Hymenoptera sphecidaenests was performed using XRF, XRD and FTIR spectroscopy. XRF data revealed the presence of SiO₂, and Al₂O₃ in appreciable quantities, while Fe₂O₃, CaO and MgO were in minor quantities. Na₂O and K₂O were however found to be present in trace amounts. Infrared spectral analysis showed that the sample is a composite of quartz, feldspar, kaolinite and montmorillonite. The dominant presence of quartz was further confirmed by PXRD which also revealed the presence of halloysites.

Keywords: Mud-wasp nest, quartz, feldspar, kaolinite, montmorillonite and halloysite.

Introduction

Hymenoptera sphecidae (mud-wasp) is a species of solitary wasps which construct nests primarily from mud, which they plaster on horizontal and vertical surfaces including walls, plant stems and others^{1,2}. These mud-wasp nests when left undisturbed for extended periods, deface the aesthetics of such buildings. When removed, the nest could be utilized gainfully instead of being thrown away as an unwanted material. Over the years, increased industrialization and urbanization has led to the disposal of hazardous substances such as heavy metals into the environment³. These industrial activities include processes of textile manufacturing, fertilizer applications, glass and ceramics. mining, electroplating, metal finishing, pulp and paper industries, petroleum and others⁴⁻⁶. Metallic elements that have a density greater than or equal to 6.0g/cm³ are regarded as heavy metals⁴. Metals that fall into this category include lead (11.34g/cm³), copper (8.95g/cm³), chromium (7.19g/cm³), cadmium (8.65g/cm³), among others⁵. The discharge of waste containing heavy metal into the environment causes serious soil and water pollution, endangering the quality of natural resources and water used for domestic applications. Heavy metals have bio-accumulating tendency and because they are also toxic in the environment, especially as most of them are carcinogenic, a problem assumed to be related to their electronic structure⁶⁻⁸ there is a need to considerably reduce their levels in industrial and municipal effluents to meet regulatory standards before final discharge into the environment.

Conventional methods used for the removal of heavy metals including floatation, chemical precipitation, solvent extraction, ion exchange, oxidation, reduction, filtration, reverse osmosis and membrane technologies⁹. These have been found to be

operational at very high cost and most times are ineffective at low concentrations of the pollutant metals¹⁰. Hence, the search for alternative metal removal techniques which are based on metal-sequestering properties of certain natural materials¹¹.

Though numerous adsorbents including commercial activated carbons have been developed and applied in chemical techniques for heavy metal removal from contaminated waste water, the high cost of these materials coupled with their failure to adsorb a range of metal ions simultaneously has been a major setback¹²; hence further research into the potential application of low-cost adsorbent materials is necessary for heavy metal removal from effluents. The use of innovative, cheap adsorbents including activated carbons generated from agricultural by-products and biosorbents has been extensively reported¹³.

The present study sought to characterize *Hymenoptera* sphecidae (mud-wasp) nests and ascertain their potential for gainful applications.

Material and Methods

Analytical grade reagents were used without further purification. Mud-wasp nest samples were collected from the walls of buildings in Makurdi metropolis, Benue State, Nigeria using plastic trowels. They were air-dried, ground with a laboratory porcelain mortar and pestle, and sieved through a 2 mm mesh. The samples were then stored in clean dry plastic sample containers and properly labelled.

Physicochemical Characterization: Determination of pH: 1.0g portion of the prepared sample was added to 100mL of distilled water in a 150mL beaker. The mixture was stirred

Vol. 3(10), 1-7, October (2013)

continuously for one hour after which it was filtered on a Whatman filter paper and the pH of the filtrate was measured by a Hanna H1 9024 pH meter.

Determination of bulk density: The tamping method described by Ahmedna $et\ al^{13}$ was employed for the determination of bulk density. In this method, 2.0g portion of mud-wasp nest was weighed and placed in a dry graduated 5mL measuring cylinder. The cylinder was then tapped until it completely occupied a minimum volume. Bulk density was then calculated using the expression in equation (1).

$$\rho_B(g/mL) = m/V_{\min} \tag{1}$$

where m = mass of mud-wasp nest, $V_{\text{min}} = \text{minimum volume}$ of mud-wasp in measuring cylinder.

Determination of Porosity: 1.0 g mud-wasp nest sample was weighed out and dispersed in 50 mL distilled water in a graduated tube. The mixture was centrifuged in a Hermle Labnet Z206A model centrifuge for 10 minutes at 5000 rpm. The resulting volume was read as $V_{\rm w}$ and recorded and porosity calculated using equation (2).

$$\alpha = V_W / V_T \tag{2}$$

Where V_W = volume of water taken

 V_{T} = volume resulting after the dispersion of mud-wasp nest.

Determination of moisture content: The moisture content was determined by the procedure given by the Association of Official Analytical Chemists (AOAC)¹⁴.

Determination of attrition: Attrition was determined as described by Toles *et al*¹⁵. In this method, 20g portion of the prepared sample was weighed and dispersed in 100mL sodium acetate-acetic acid buffer of pH 4.0 prepared by mixing aqueous 0.1 M CH₃COONa solution and commercial CH₃COOH in the ratio 1:5.5 in a 250mL glass beaker and left to stand for 2 h at room temperature. The suspension was filtered through a 2mm sieve and the amount of mud-wasp nest sample retained was quantitatively transferred into a pre-weighed aluminium pan and dried in an oven at 105°C. Attrition was calculated as the ratio of mass loss of the mud-wasp nest to the initial mass expressed as a percentage from the equation(3).

% Attrition =
$$\frac{m_i - m_f}{m_i} \times 100$$
 (3)

Where mi = initial mass of mud-wasp nest, mf = final mass of mud-wasp nest.

Determination of iodine adsorption number (IAN): The iodine adsorption number was determined by the procedure described by Okieimen *et al*¹⁶. In this method, 0.5g portion of mud-wasp nest was slurred with excess aqueous 0.05M iodine solution in a 250mL glass beaker. The content was then swirled vigorously for 10minutes and the mixture filtered through a

funnel impregnated with glass wool. A portion of the filtrate was back-titrated with a standard solution of thiosulphate. The mass (mg) of iodine consumed per gram of mud-wasp nest constitute the iodine number.

Determination of surface area of nest material: Surface area (S.A.) was calculated as the inverse of the iodine number¹⁶.

$$S. A. = \frac{1}{IAN}$$
 (4)

Determination of titrable surface charge: The Boehm titration described by Van-Winkle¹⁷ was employed in the determination of titrable surface functional groups. 1.0g portion of mud-wasp nest was suspended in 50mL aqueous 0.1 M NaOH solution with occasional stirring for 12 h. The slurry was then filtered through glass wool impregnated in the stem of a plastic funnel. 10mL aliquots of the filtrate were added to 15mL of standard aqueous 0.1M HCl solution and the resulting solution was back-titrated with standard aqueous 0.1M NaOH solution. The volume of NaOH required to neutralize the sample was converted into titrable negative surface charge by expressing the result as millimoles H⁺ ions consumed by excess OH⁻ ions per gram of sample.

Determination of particle size distribution: Particle size distribution was determined by the hydrometer method¹⁸. 50 g of 2 mm sieved mud-wasp nest sample was weighed into a 250 mL glass beaker. 100 mL aqueous sodium hexameta phosphate (calgon) solution was added into the beaker followed by 100 mL of distilled water. The suspension obtained was stirred with a glass rod and left to stand for 30 minutes with occasional stirring. The suspension was transferred completely into a 100 mL measuring cylinder and made up to the mark with distilled water. Before taking hydrometer readings, the suspension was thoroughly mixed with a glass rod making sure that the sediments at the bottom of the cylinder were thoroughly disturbed. A soil hydrometer was then lowered gently into the suspension and readings taken after 40 seconds. The suspension was then allowed to stand for 2 h and hydrometer readings taken. The temperature of the suspension was also measured with a thermometer and recorded appropriately. A blank cylinder was prepared with 50% calgon solution made up to the 1L mark with distilled water. Blank hydrometer readings at 40 seconds and 2 hours were obtained as previously described. Particle sizes were then calculated from equations (5, 6, 7, 8)

$$C(g/L) = R - R_L + (0.36 T)$$
 (5)

where C = corrected hydrometer reading (g /L), R (g /L) = Hydrometer reading in the suspension, R_L (g /L) = Blank hydrometer reading, T = temperature (20°C for this research))

% Clay =
$$\frac{corrected 2h \ reading-blank \ reading}{weight \ of \ soil \ used} \times 100$$
 (6)

% Silt =
$$\frac{\text{corrected 40 s reading-blank reading}}{\text{weight of soil used}} \times 100 - \% \text{ clay}$$
 (7)

% Sand =
$$100 - (\% \text{ clay} + \% \text{ silt})$$
 (8)

Determination of organic matter: The classical Walkley-Black rapid oxidation method was used for the determination of organic matter content¹⁹. 0.1g of mud-wasp nest was weighed out and transferred to a 250 mL conical flask. Using a pipette, 10 mL aqueous 0.167 M K₂Cr₂O₇ solution was added to the solution in the conical flask and swirled gently. 20mL portion of concentrated H₂SO₄ was then added and the mixture swirled gently to mix. Excessive swirling was avoided to prevent organic particles from adhering to the sides of the flask out of the solution. The contents of the flask were allowed to stand for 30minutes by placing it on a sheet of asbestos to avoid rapid heat loss. The content of the flask was then titrated against 0.5M ferrous ammonium sulphate solution (FAS). As end point was approached, the colour of the solution gradually changed to greenish depending on the amount of unreacted dichromate. At this stage, FAS was added drop-wisely until a sharp colour change of wine red was attained to mark the end point, viewed against a white -tile background. Blank titrations were performed in the same manner without the mud-wasp nest. Percent carbon and percent organic matter were calculated using equations (9) and (10).

% oxidizable organic carbon,

$$\%C = \frac{(B-S) \times \text{molar conc. of } Fe^{2+} \times 12}{\text{mass of sample } (g) \times 4000} \times 100$$
 (9)

where $B = \text{volume (mL) of Fe}^{2+}$ solution used to titrate blank, S= volume (mL) of Fe²⁺ solution used to titrate sample, $\frac{12}{4000}$ = milliequivalent weight (g) of carbon

Oxidizable organic carbon was converted to total carbon by dividing by 0.77^{19} % Organic matter = %C x $\frac{1.72}{0.58}$

% Organic matter = %C x
$$\frac{1.72}{0.58}$$
 (10)

Spectrophotometric analysis of Hymenoptera sphecidae (mud-wasp) nest: X-ray fluorescence (XRF) analysis: The fine powder material was pelletized and the solid pellet was then fed into a thermo 9900 intellipower x-ray fluorescence spectrophotometer for analysis.

Fourier transforms infrared (FTIR) analysis: FTIR data were collected on a Shimadzu FTIR-8400s Fourier Transform Infrared Spectrophotometer. Measurements were carried out in the range of 4000-200 cm⁻¹.

Powder x-ray diffraction (PXRD) analysis: Powder x-ray diffraction data of Hymenoptera sphecidae nest were obtained on a Bruker D8 diffractometer using Cu k_{α} (1.5406Å) radiation, over 2θ-range between 5°- 90° using a step size of 0.014° and step time of 0.2s.

Results and Discussion

Physicochemical characteristics of Hymenoptera sphecidae (mud-wasp) nest: Results of the physicochemical properties of Hymenoptera sphecidae are presented in table 1.

Characteristics of Hymenoptera sphecidae(mud-wasp) nest

Characteristics of Hymenoptera spineetaate (mad wasp) nest						
Parameter	Value					
pН	6.54 ± 0.02					
Abrasion resistance (%)	95.10 ± 0.10					
Bulk density (g/cm ³)	1.27 ± 0.02					
Porosity (mg/g)	0.96 ± 0.01					
Moisture content (%)	1.29 ± 0.10					
Total surface charge (mmolH ⁺ eq/g)	1.35 ± 0.02					
Iodine adsorption number (IAN) (mmol/g)	0.16 ± 0.20					
Total surface area (g/mgI ₂)	6.33 ± 1.60					
Organic matter (%)	4.77 ± 1.60					
Particle size distribution :						
% clay	10.96 ± 0.50					
% silt	15.52 ± 3.37					
% sand	73.52 ± 2.83					
Cation exchange capacity (meq/100g)	0.15 ± 0.01					

The pH value of 6.54±0.02for Hymenoptera sphecidae (mudwasp) nest indicated that the material is acidic. Generally, for pH-sensitive adsorbates, adsorbent pH in the range of 6-8 is acceptable for most adsorption applications in aqueous phase media¹⁶⁻²⁰. On the other hand, adsorbents, particularly those derived from agricultural residues have indicated an optimum metal ion uptake level at pH > 4.0^{-15} . Previous studies^{15,16} have shown that adsorbents are capable of increasing or decreasing the pH of slurries to levels that are out of range for metal ion adsorption.

The percent abrasion resistance also known as attrition is an indicator of the mechanical strength of an adsorbent for aqueous phase applications. It describes the ability of the adsorbent to, not only maintain its physical integrity during the adsorption process but also its ability to withstand frictional forces imposed by back washing 16. Previous investigations have revealed that for an adsorbent, particularly the low-cost ones to be considered for economic purposes, it should possess not only impressive adsorption properties, but also a high resistance to abrasive forces in batch and column applications¹⁶. Low abrasion resistance leads to loss of adsorbent particle integrity and formation of dust particles, resulting in the reduction in the rate of filtration, and in the amount of the adsorbent¹⁶. Percent attrition for mud-wasp nest determined was 95.10±0.10%, a value interpretive of good mechanical strength, implying its potential for low degradability during handling.

Bulk density is one of the parameters that describe the quality of an adsorbent. It predicts the filterability of an adsorbent. High values of bulk density portend good quality adsorbents²¹. The investigated mud-wasp nest has a bulk density value of Vol. 3(10), 1-7, October (2013)

Res. J. Chem. Sci.

1.27±0.02 g/cm³, a comparatively high value with regard to other adsorbents reported in literature^{2,15,16}.

Porosity of an adsorbent is a measure of the void in the adsorbent; and the greater the porosity of given adsorbent, the greater would be the relative size of molecules it can adsorb onto its crystal structure. Thus, the ion exchange or adsorption characteristics of adsorbent are determined also by the size and geometry of its pore spaces. In this study, mud-wasp nest has porosity of 0.96±0.01, which shows great promise for adsorption applications.

The amount of moisture present in the mud-wasp nest was determined to be 1.29±0.10%, a value far lower than 50.00% upper limit for adsorbents²². This implies that the mud-wasp nest can be stored for long periods without significant microbial activity being observed.

The adsorption capacity of an adsorbent is influenced by the presence of surface functional groups ¹⁶. Surface functional groups that dominate adsorption behaviour of an adsorbent include carbonyls, carboxyls, phenolics, lactones, etc. ^{16,22,23}. These impart not only polarity onto the surface of the adsorbent, enhancing adsorption of charged species, but also influence possible mechanisms for covalent bonding and surface catalysis. The total surface charge for mud-wasp nest determined was 1.35±0.02mmolH⁺eq/g, which is comparable with values that have been reported for some adsorbents including commercial grade carbons²².

Iodine adsorption number (IAN) is a fundamental parameter usually used to characterize the performance of an adsorbent²⁴. IAN value for mud-wasp nest was 0.16±0.20mmol/g, which is comparatively lower than reported for other adsorbents²². This implies that further treatment of mud-wasp nest will be desirable to enhance its adsorptive capacity.

Surface area of an adsorbent usually measures the extent of pore surface developed within the matrix of the adsorbent. Its value indicates the functionality of an adsorbent based on the principle that the greater the surface area, the higher the number of available adsorption sites¹⁶. A large surface area is therefore, a necessary requirement for a good adsorbent. When the internal pore structure of a material is highly developed the surface area is large, presenting the material as adsorbent and imparting in it the ability to adsorb gases and vapors from gas streams and dissolved/ dispersed substances from liquid media²⁴⁻²⁵. The investigated mud-wasp nest gave the total surface area value of 6.33±1.60g/mgI₂, which compares with values reported for other effective adsorbents^{16,19}.

CEC of soil (meqH⁺/100g) is a measure of the amount of sites on the soil surface that can retain cations through electrostatic force means. Cations retained by electrostatic means can easily be exchanged with other cations present and available in the soil environment²⁶. Sites for cation exchange are found mainly on clay and organic matter (OM) surfaces. In this investigation, the CEC for mud-wasp nest was 0.15±0.01meq H⁺/100g. Normal CEC ranges in soils have been reported to be from 3meq H⁺/100g for sandy soils low in organic matter to >25meq H⁺/100g for soils high in certain types of clay or OM²⁶. The cation exchange capacities of alkaline soils are commonly higher than those of acidic soils with comparable soil textures. This is due to two primary factors namely, the high CEC associated with the constant charges on 2:1 type clays that are most common in alkaline soils; and the even higher CEC resulting from the pH dependent charges on the humus colloids at these pH levels²⁶.

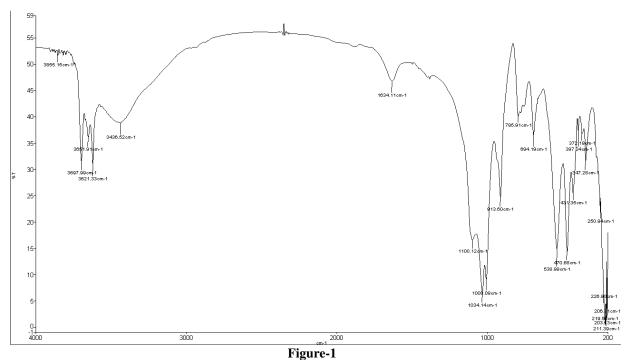
Mineral composition of *Hymenoptera sphecidae* (mud-wasp) nest: X-ray fluorescence analysis: XRF analysis of *Hymenoptera sphecidae* nest revealed that silica and alumina were present in dominant quantities of 26.5995%, and 10.5095% respectively (table 2). Calcium oxide, magnesium oxide and iron (III) oxide recorded values of 3.406%, 3.7015% and 2.3715% respectively. Trace amounts of sodium and potassium oxides were recorded as 0.9675% and0.3010 % respectively.

Fourier Transform Infrared (FTIR) analysis: Heating the KBr in the oven at 120°C was to prevent the broad spectral peak due to free OH groups from seriously affecting the interpretation on the bound hydroxyls associated, with any of the minerals²⁰. Absorption of infrared energy by a mineral is associated with the vibrational and rotational motion of molecules within it²⁷. The infrared spectrum obtained for *Hymenoptera sphecidae* nests is presented in figure 1. Quartz, feldspar, kaolinite and montmorillonite were identified from the absorption frequencies in the spectrum.

Quartz is one of the world's most abundant minerals and an important component in the earth's crust. It is primarily silicon dioxide, SiO₂²⁷. Strong IR absorption bands appeared at 795.91cm⁻¹ and 694.17cm⁻¹ suggesting the presence of quartz in the sample²⁷. The absorption band observed at 694cm⁻¹ was assigned to Si-O symmetrical bending vibrations^{20,27} while the band at 795.91cm⁻¹ indicated the presence of Si-O symmetrical stretching vibrations.

Table-2
Mineral composition of hymenoptera sphecidae (mud - wasp) nest

Compound	SiO ₂	$\hat{Al_2O_3}$	Fe ₂ O ₃	CaO	MgO	Na ₂ O	K ₂ O
% w/w	26.5995	10.5095	2.3715	3.4060	3.7015	0.9675	0.3010



Infrared spectrum for hymenoptera sphecidae (mud-wasp) nest

Feldspar is an important group of rock-forming minerals with a general formula WZ_4O_8 whereWmay be Na, K, Ca or Ba while Z is Si and/ or Al, the Si:Al ratio ranging from 3:1 to 1:1²⁰. Feldspar group of minerals have been analyzed by IR technique and reported²⁰. The sharp IR absorption peak appearing at 538.99cm⁻¹ was assigned to feldspar and is due to Si-O asymmetric bending vibrations and Si-O-Al stretching vibrations, which often pertain to the 540-535cm⁻¹ region^{20,27} of the IR spectrum.

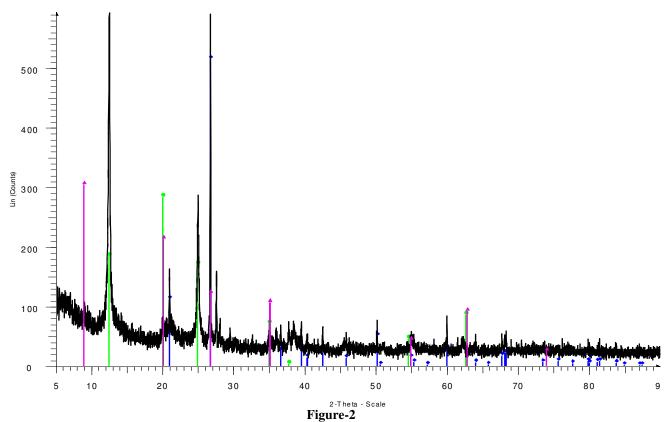
Kaolin is a mineral with chemical composition $Al_2Si_2O_5(OH)_4It$ is a layered silicate mineral with one tetrahedral sheet linked through oxygen molecules to one octahedral sheet of alumina²⁰. Intense IR absorption bands appearing at $3697.99cm^{-1}$, $3661.91cm^{-1}$, $3621.33cm^{-1}$, $1034.14cm^{-1}$, $1100.14cm^{-1}$, 1008.09 cm⁻¹ and 470.68 cm⁻¹ in the spectrum indicated the presence of kaolinite^{20,27}. The strong absorbance at $3697.15cm^{-1}$ was assigned to inner surface OH stretching vibrations, while the band observed at $3621.33cm^{-1}$ was attributed to inner OH stretching vibrations indicating hydroxyl linkage^{20,27}. The absorbance at $1100.12cm^{-1}$, 1008.09 cm⁻¹ and $1034.14cm^{-1}$ were indicative of anti-symmetric Si-O-Si stretching vibrations²⁷, while the band at $470.68cm^{-1}$ was due to Si-O and Si-O-Fe stretching vibrations²⁸.

Montmorillonite is one of the very soft and tenderphyllosilicate minerals. It is primarily hydrated magnesium calcium aluminium silicate (Mg, Ca) O.Al₂O₃.5SiO₂.nH₂O with *n* ranging from 5 to 7. From the spectrum in figure 1, the presence of the broad IR absorption bands at 3436.52cm⁻¹ and 1634.11cm⁻¹ indicated the presence of montmorillonite^{20,27}. The

band at 3436.52cm⁻¹ was assigned to H-O-H stretching of absorbed water molecules²⁸, while the strong band at 1634.11cm⁻¹ suggested OH deformation of water²⁰.

Powder X-ray diffraction (PXRD) analysis: Qualitative mineralogy of *Hymenoptera sphecidae*nests was further tested by PXRD analysis. The x-ray diffractogram for the sample is presented in figure 2. The PXRD pattern of the sample when compared against the ICCD database for the theoretical phases shows that the dominant mineral present in mud-wasp nest is quartz (92.71%, pattern number = 01-087-2096), while halloysite7A- Al₂Si₂O₅(OH)₄ (pattern number = 00-029-1487) and halloysite10A-Al₂Si₂O₅(OH)₄.2H₂O (pattern number = 00-029-1489) are present as 47.70% and 13.86% respectively. Halloysite is a 1:1 alumino silicate clay mineral and like kaolinite, it has a single layer structure. The primary difference between halloysite and kaolinite is their structure. Halloysite has a fibrous structure while kaolinite has a platestructure²⁹.

Because of the fibrous habit of halloysite, crystals are not oriented in basal parallel fashion. This results in weak 001 (hkl) reflections that are generally broad and strong non-basal reflections between $20^{\circ}-30^{\circ}~2\theta^{29}$; as a result, halloysite has a diffraction pattern with broad 001 and strong asymmetrical hkl reflections²⁸. Primary and secondary quartz were observed in the peak locations at 26.5° (d-space =4.251Å) and 20.9° (d-space = 4.251Å respectively²⁹, while tertiary and quarternary quartz were identified at peak locations of 50.0-50.9° (d-space =1.817Å) and 60.0° (d-space =1.541Å) respectively²⁹.



Powder X-ray Diffraction (PXRD) spectra for Hymenoptera sphecidae (mud-wasp) nest

Conclusion

Physicochemical characteristics of *Hymenoptera sphecidae* (mud-wasp) nest were investigated andvalues obtained compared favorably with those of other adsorbents previously studied by other researchers. XRF data revealed that SiO₂, and Al₂O₃ were the major minerals present, while Fe₂O₃, CaO and MgO were present in minor quantities. Na₂O and K₂O were however found to be present in trace amounts. From FTIR absorption peaks obtained, the constituent minerals identified were quartz, feldspar, kaolinite and montmorillonite. The dominant presence of quartz was further confirmed by PXRD which also revealed the presence of halloysites. Therefore, *Hymenoptera sphecidae*nest is a composite material, which may have very promising application in adsorption studies.

Acknowledgements

We appreciate the quality control unit of Benue Cement Company Ltd for running the XRF analysis for this study. The FTIR and PXRD analyses were done by the research laboratory at the Chemistry Department of Loughborough University, Loughborough United Kingdom. The assistance of the technical staff headed by Mr G. H. Atoo at the Department of Chemistry, Benue State University, Makurdi-Nigeria is very highly appreciated.

References

- 1. Huntera J.M., Insect clay geography in Sierra Leone, Journal of Cultural Geography, 4(2), 13 (1984)
- 2. University of Pretoria publication, http://www.University of Pretoria pubs.com, Retrieved February 16th 2011 (2011)
- 3. Okparaeke O.C., Agha I.I. and Ejikeme P.M., Removal of Cu(II) and Hg(II)ionsfrom simulated waste water by adsorption on tounactivated/activated carbon from *Brachtystagea Eurycoma* seed pods, Intraparticulate diffusivitysorption studies, *Jour.Chem. Soc.*, 35(1), 94-98 (2010)
- **4.** Horsfall M. and Ayebaemi I.S., (2005), Equilibrium sorption study of Al³⁺, Co²⁺ and Ag⁺ in aqueous solution by fluted pumpkin (*Telfariaoccidendalis* HOOKf) waste biomass, *Acta. Chim. Slov.*, **52**, 174-181 (**2010**)
- 5. Njoku V.O., Oguzie E.E., Obi C., Bello O.S. and Ayuk A.A., Adsorption of copper (II) and lead (II) from aqueous solutions onto Nigerian natural clay, *Australian Journal of Basic and Applied Science*, 5(5), 346-353 (2011)
- **6.** Keskinkan O., Goksu M.Z.L., Basibuyuk M. and Forster C.F., Heavy metal adsorption characteristics of a

- submerged aquatic plant (Myriophyllumspicatum), Process Biochem., 39, 179-183 (2003)
- 7. Okonkwo A.E. and Anwasi S., Modelling of copper and zinc adsorption from aqueous solutions by *Thithoniadiversifolia*, *Journal of Chemical Society of Nigeria*, 3(1), 66-72 (2010)
- **8.** Kelth L.H. and Telliard W.A., Priority pollutants, *Environmental Science and Technology*, **13**, 416 (**1979**)
- **9.** Igwe J.C., Ekeghe E.M.N. and Abia A.A., Binding of Cu²⁺, Cd²⁺, and Hg²⁺ions from aqueous solutions onto thiolated and carboxymethylated sawdust, *International Journal of Chemistry*, **16(3)**, 121-128 (**2006**)
- **10.** Abechi S.E. and Gimba C.E., Adsorption of cadmium from aqueous solution by activated carbon prepared from sawdust and walnut shell, *Journal of Chemical Society of Nigeria*, **35(1)**, 1-4 (**2010**)
- **11.** FAO/WHO, Evaluation of certain food additives and contaminants mercury, lead & cadmium.Food and agricultural organization/World health organization 16th report, Rome, 84 (**1972**)
- **12.** Chukwuma C.S., Environmental lead exposure in Africa, *Am. Bio.*, **26(6)**, 399-403 (**1997**)
- **13.** Ahmedna M.S., Clarke S.J., Rao R.M., Marshall W.E. and John M.M., Use of filtration and buffers in raw sugar colour measurements, *Jour. Sci. Food Agric.*, **75**, 109-116 (**1997**)
- **14.** A.O.A.C., *Official methods of analysis*, 13th edition. Washington D.C, 1-7 (**1990**)
- **15.** Toles C.A., Marshall W.E., Johns M.M., Wartelle L.A. and McAloon A., Acid activated carbons from almond shells: physical chemical & adsorptive properties & estimated cost of production, *Bioresources Techno.*, **71**, 87-92 (**2000**)
- **16.** Okieimen F.E., Okieimen C.O. and Wuana R.A., Preparation and characterization of activated carbon from rice husks, *Jour. Chem. Soc. Nig.*, **32(1)**, 126-136 (**2007**)
- **17.** Boehm H.P., Some aspects of the surface chemistry of carbon blacks and other carbons, *Carbon*, **3(5)**, 759-760 (**1994**)
- **18.** Bouyoucos G.J., Improved hydrometer method for making particle size analysis of soils, *Agron. Jour.*, **54**, 464–465 (**1962**)

- **19.** Schumacher B.A., Methods for the determination of total organic carbon (TOC) in soils and sediments, USEPA NCEA C-1282 (**2002**)
- **20.** Ravisankar R., Kiruba S., Chandrasekaran A., Senthikumar G. And Maheswaran C., Analysis of ancient potteries of Tamilnadu, India by spectroscopic techniques, *Ind. Jour. Sci. Tech.*, **3(8)**, 858-862 (**2010**)
- **21.** Hasany S.M. and Chaudhary M.H., Sorption potential of Hare river sand for the removal of Antimony from acidic aqueous solution, *Appl. Rad. Isot.*, **47**, 467-471 (**1996**)
- **22.** Adejo S.O., Wuana R.A., Leave E.T. and Angba O.M., Evaluation of physicochemical and adsorptive properties of adsorbents prepared from guinea corn (Sorghum bicolor) husks, *Nig. Jour. Pure App. Sci.*, **1**, 1-9 (**2008**)
- **23.** Wartelle L.H. and Marshall W.E., Nutshells as granular activated carbons: physical and adsorptive properties, *Jour. Chem. Tech. Biotech.*, **76**, 451-455 (**2001**)
- **24.** Weber T.W. and Chakkravorti P., Pore and solid diffusion models for fixed bed adsorbents, *A.I.Chem. Jour.*, **20**, 228 (1974)
- **25.** Okafor J.O. and Aneke N.A.G., Evaluation of sorption capacity of locally developed activated carbon using methylene blue number method, *Chem. Class Jour.*, 124-127 (**2007**)
- **26.** Gillman G.P. and Sumpter E.A., Modification to the compulsive exchange method for measuring exchange characteristics of soils, *Aust. Jour. Soil Res.*, **24**, 66-68 (1986)
- 27. Ravisankar R., Senthilkumar G., Kruba S., Chandrasekaran A. and Prakash J.P., Mineral analysis of coastal sediment of Tuna Gujarat, *India, Ind. Jour. Sci. Tech.*, 3(7), 774-780 (2010)
- 28. Nayak P.S. and Singh B.K., Instrument characterization of clay by XRF, XRD & FTIR, *Bull. Mater. Sci.*, 30(3), 235-238 (2007)
- **29.** Duncan W.S., Ledger E.B. and Whitehead V.S. (n.d) X-ray diffraction verification of Aviris clay mineral identification, Summitville area, Southeast Colarado, Nocogdoches, SFASA, 1-9 (1998)